

Putative role of 5-HT_{2B} receptors in migraine pathophysiology

Cephalalgia

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Abstract

Objective: In this review we attempt to characterize the acute and chronic role of 5-HT_{2B} receptors with regard to meningeal nociception in animal experiments and clinical data targeting migraine therapy.

Background: Migraine is a common disabling neurovascular primary headache disease, the pathomechanism of which is still unclear. Serotonin (5-HT) and its receptors might play an important role in some aspects of migraine pathogenesis. The ability of the unselective 5-HT_{2B} receptor agonist m-chlorophenylpiperazine to induce migraine attacks in migraine sufferers, the high affinity of prophylactic antimigraine drugs to this receptor and its expression in migraine-relevant structures like the dura mater argue for a role of 5-HT_{2B} receptors in the pathogenesis of migraine attacks.

Methods: For this review, the relevant databases such as PubMed, MEDLINE[®], Cochrane Library and EMBASE, respectively, were searched to December 2015 using the keywords “migraine, 5-HT₂, trigeminal, neurogenic inflammation, nitric oxide, nitroxyl, vasodilatation, plasma protein extravasation” and combinations thereof.

Conclusion: Our literature review suggests an important role of 5-HT_{2B} receptor activation in meningeal nociception and the generation of migraine pain.

Keywords

5-HT_{2B} receptor, vasodilatation, NO dependent, CGRP, plasma protein extravasation

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Introduction

Serotonin (5-HT) and its receptors play an important role in migraine research. The concentration of the 5-HT metabolite, 5-hydroxyindole acetic acid (5-HIAA), has been found increased in urinary excretion during migraine attacks and could thus serve as a biomarker of migraine (1). 5-HT is released from platelets by compounds such as fenfluramine, while reserpine or 5-HT reuptake inhibitors (zimeldine and femoxetine) may increase the frequency of migraine attacks (2,3). During migraine attacks the 5-HT concentration is decreased in platelets and subsequently increased in the blood plasma (4–6). Brewerton et al. reported that administration of m-chlorophenylpiperazine (mCPP), a metabolite of the antidepressant tradozone and nefazodone, with different affinities to 5-HT_{2B/2C} receptors and other serotonin, adrenergic, dopamine and muscarine cholinergic receptors, leads to delayed “typical migraine headache” in migraineurs and unspecific headaches in healthy individuals (7). Experimental data show that particularly 5-HT_{2B} receptors play a role in functions that may be

associated with the pathophysiology of headaches. Acute activation of 5-HT_{2B} receptors by mCPP led to nitric oxide (NO)-dependent plasma protein extravasation (PPE) in the dura mater of guinea pigs (8,9) and activation of neurons in the trigeminal nucleus caudalis (TNC) of rats (10). Therefore it has been assumed that meningeal 5-HT_{2B} receptors are involved in early steps of migraine pathogenesis (11). Clinical evidence for a role of 5-HT₂-receptors in migraine is provided by the fact that various prophylactic agents (methysergide or pizotifen) are 5-HT₂ receptors antagonists.

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Structure and localization of 5-HT_{2B} receptors

The 5-HT_{2B} receptor was first described by Vane in the rat gastric fundus, where it is responsible for the contraction of the longitudinal muscle cell layer (12). In 1992, 5-HT_{2B} receptors were first cloned from rats and mice (13,14) followed by cloning of the human 5-HT_{2B} receptor (15,16). The receptor protein consists of 481 amino acids (16) and has a 45% homology with 5-HT_{2A} and a 42% homology with the 5-HT_{2C} receptor protein (17). The occurrence of the 5-HT_{2B} receptor is ubiquitous. It occurs with a particularly high rate in liver and kidney, while a low expression rate was detected in pancreas and spleen (18). Likewise it is expressed in lung arterial endothelial cells, where it is responsible for the development of pulmonary hypertension (19). In the rat brain 5-HT_{2B} receptors are slightly expressed in neurons located in the cerebellum, the posterior hypothalamus, the lateral septum and the medial amygdala but these data, which are derived from immunohistochemical staining (IHS) (20), are considered controversial. Other studies using IHS and molecular biological methods showed expression in motoneurons (21), in rat spinal cord and in dorsal root ganglion (DRG) (22–24). The subcellular localization of the receptor in blood vessels is not evident from the literature; however, it has been suggested that it is localized on the luminal side of vascular endothelial cells (11).

An important function in the context of the 5-HT_{2B} receptor is the activation of nitric monoxide synthase (NOS), which promotes cleavage of the guanidino group from the amino acid arginine and other intermediate steps of NO synthesis (25). According to experiments in cell cultures, NOS seems to be coupled to the 5-HT_{2B} receptor through a PDZ-domain of the c-terminus (26). The link between the receptor and NOS in vivo is still unclear, and direct measurements in vivo are difficult because of the low number of 5-HT_{2B}-positive cells (27).

Location of the receptor in structures relevant for migraine

Schmuck et al. succeeded in preparing RNA transcripts from migraine-relevant structures such as the dura mater (11). The receptor in the dura mater could be detected in endothelial cells and it was also found weakly expressed in smooth muscle cells. Lin et al. detected messenger RNA (mRNA) of the 5-HT_{2B} receptor by in situ hybridization in trigeminal ganglion (TG) neurons of mice. A specific function of the receptor in the TG is not yet known but it can be assumed to be transported along peripheral and central processes of trigeminal afferents (23).

The acute activation of 5-HT_{2B} receptors leads to neurogenic inflammation

Components of meningeal neurogenic inflammation, particularly vasodilatation and plasma protein extravasation (PPE) induced by the release of neuropeptides from primary sensory nerve terminals, have frequently been observed in animal experiments as parameters of meningeal nociception (28). Although these changes are not directly responsible for the headaches, they may reflect peripheral pathophysiological events associated with migraine pain. In a study on guinea pigs, acute intravenous administration of 1 µg/kg mCPP induced PPE in dura mater (determined with intravenous Evans blue application), which could be inhibited by selective 5-HT_{2B} receptor antagonists (LY53857, LY215840; both 10 µg/kg), whereas selective inhibition of the 5-HT_{2C}-receptor (using the antagonist LY310898) had no influence on the PPE (9). It should be mentioned that, in addition to different 5-HT receptors, mCPP interacts also with other neurotransmitter receptors such as adrenergic, dopamine and muscarine cholinergic receptors (for overview see Hamik and Peroutka (29) and Martin and Martin (10)).

Schmitz et al. showed similar results with the same preparation using a new substance (BF-1) for blocking PPE. BF-1 has very high affinity for the 5-HT_{2B} receptor ($pki = 8.63 \pm 0.24$, SD) with only minor affinity to the 5-HT_{2C} receptor ($pki = 7.64$) and the histamine 1 (H1) receptor ($pki = 7.81$). According to the authors these low affinities for the 5-HT_{2C} receptor and the H1 receptor preclude unspecific effects, which appeared to be more prominent with older substances, e.g. pimehixene (1-methyl-4-(9H-thioxanthen-9-ylidene) piperidine ($pki = 10.14$ for H1 receptors and 8.42 for 5-HT_{2C} receptors) (8). The inhibition of 5-HT_{2B} receptors with the compound RS-127445, a selective, high-affinity ($pki = 9.5$), orally bioavailable 5-HT_{2B} receptor antagonist (30), was shown to inhibit the mCPP-induced PPE as well as c-fos expression in the rat trigeminal nucleus caudalis (TNC) evoked by capsaicin (31). Schmuck et al. has also shown that the activation of 5-HT_{2B} receptors by the unspecific agonist dihydroergotamine (DHE) (10^{-9} to 10^{-6} mol/l) causes vasodilatation of isolated cerebral arteries in the pig. The DHE-induced relaxation could be blocked by pizotifen (10^{-6} mol/l), a prophylactic antimigraine drug (11). As a limitation for this association it should be mentioned that pizotifen also has an antihistamine (H₁-receptor blocking) and weak anticholinergic action, which could be involved in the prophylactic effect (32–34). However, taking together the above results, it seems most likely that it is the 5-HT_{2B} receptor that is involved in formation of the major components of neurogenic

inflammation, vasodilatation and PPE evoked by mCPP. The neurogenic inflammation is induced by the release of substance P, which is mainly responsible for the PPE, and calcitonin gene-related peptide (CGRP), which mediates vasodilatation. Substance P binds to the neurokinin receptor 1 (NK-1), which is localized on endothelial cells, and thereby induces the formation of gaps in the endothelium (ca. 0.5–1.5 μm) allowing plasma proteins to diffuse into the perivascular tissue, e.g. demonstrated in the rat trachea (35). PPE has been studied extensively in different tissues including the dura mater (36). In an electron microscopic study of dural blood vessels, Dimitriadou found that the electrical-induced PPE is not caused by an increased number of endothelial gaps but rather by an increased number of pinocytotic vesicles (37). Hunfeld et al. observed an mCPP-induced PPE that was associated with increased transcellular transport of dissolved substances (e.g. horseradish peroxidase, HRP) in endothelial cells with no damage of fenestrae or tight junction integrity in mice dura mater (38). Other studies showed both transcellular and paracellular transport through the endothelial junction and clefts (39). However, the precise mechanism of PPE in the dura mater and whether it is important for the emergence of migraine remains questionable, because blockade of the PPE via NK-1 receptor antagonists turned out to be ineffective in migraine treatment (40,41).

The extent of vasodilatation is routinely measured by parameters such as an increased vessel diameter or increased blood flow (42,43). Furthermore, substance P and CGRP released from meningeal afferents may be involved in the activation and degranulation of dural mast cells, which may express receptors for both neuropeptides (44). Through the release of pro-inflammatory substances such as histamine and cytokines from mast cells, the neurogenic inflammation of the dura mater may be aggravated (45).

Massive mast cell degranulation (by compound 48/80) has been shown to activate primary meningeal afferents and second-order neurons in the TNC (46). However, it is questionable if the neuropeptides as weak mast cell activators can induce such an effect, and it is unclear which substances released from mast cells are capable of activating primary afferents. One candidate, histamine, activated only a very small proportion of meningeal afferents in an *in vitro* study (47). In experimental and clinical studies administration of histamine caused dilatation of cerebral arteries and induced typical migraine headache, which could be inhibited by blockade of H1 receptors (48,49). However, inhibition of H1 receptors blocked only the “histamine-induced headache” but was ineffective in prophylactic migraine treatment (4,50). This may support the view that 5-HT, which is another substance concentrated in mast cell granules, may be more

important as a “natural player” in the generation of migraine attacks (11).

In summary, the observations that 5-HT_{2B} receptor agonists like mCPP induce plasma extravasation as the main element of neurogenic inflammation in the dura mater and increase c-fos expression in the TNC argue for a role of 5-HT_{2B} receptors in events associated with the generation of trigeminal activity and possibly migraine pain (9,10).

5-HT_{2B} receptor-induced neurogenic inflammation depends on NO synthesis

Kalkman postulated in 1994 that activation of “5-HT_{2C}-like” receptors can provoke migraine attacks because the activation of these receptors leads to NO release, which may be a key event in triggering migraine (2). Fozard already postulated this hypothesis as early as 1975 but was not sure which of the 5-HT receptors can induce the release of NO. It should be noted also that the activation of other receptors, e.g. H1 receptors, leads to NO release (51). Recent studies focused on elucidating the role of NO and its specific actions. The peripheral activation of 5-HT_{2B} receptor leads to formation of NO (11). In a study with acute 5-HT_{2B} receptor activation, administration of the NO synthase inhibitor L-NAME caused inhibition of dural PPE in the guinea pig (9). NO is known to activate the soluble guanylate cyclase (sGC) in smooth vascular muscle cells resulting in an increase in cyclic guanosyl monophosphate (cGMP) (52). By this way NO causes relaxation of smooth muscle resulting in vasodilatation of arterial blood vessels and increased blood flow (53). Recent experiments indicate alternative, possibly even more potent, vasodilatory mechanisms. In the presence of hydrogen sulfide (H₂S), a product of the condensation of cysteine with homocysteine to cystathionine, catalyzed by enzymes like cystathionine β -synthase (CBS) (54), NO can be metabolized to nitroxyl (HNO). HNO is a potent agonist of TRPA1 receptor channels, which upon their opening induce an influx of Ca²⁺ (55,56). In TRPA1 expressing peptidergic sensory neurons, this mechanism is mainly responsible for the release of neuropeptides like CGRP (56). CGRP is regarded as the most potent vasodilator of intracranial arteries (57), hence this mechanism caused strong vasodilatation and blood flow increase in the rat cranial dura mater (58).

In the TG, NO may be involved in a neuron-glia crosstalk. In TG cell cultures, NO donors caused CGRP promoter activity and secretion (59). Conversely, CGRP treatment increased glial iNOS expression and NO release from TG satellite cells (60). This could lead to a vicious circle if CGRP-releasing neurons are surrounded by NO-producing satellite cells; however, it is yet uncertain if this crosstalk takes place in the intact TG *in vivo*. Communication between CGRP-releasing and NO-

producing neurons in the TG also seems possible. Glycerol trinitrate (nitroglycerin, GTN), a substance directly activating the sGC (61), which is long known to induce vascular headaches such as delayed migraine attacks in migraineurs (62), caused upregulation of CGRP, CGRP receptor components and neuronal NO synthase (nNOS) in rat TG neurons (63,64).

Similar signaling mechanisms may take place in the superficial laminae of the TNC, where the central terminals of nociceptive trigeminal afferents synapse onto second-order neurons. CGRP released from such terminals acts as a neuromodulator facilitating synaptic transmission (65). Multiple neurons in all spinal layers seem to express nNOS (66) and potentially produce NO as a “retrograde transmitter” that facilitates neurotransmitter release from presynaptic terminals (67).

Taken together, several lines of evidence are suggestive of 5-HT_{2B} receptor-dependent NO production, which may be involved in the pathogenesis of nociceptive processes of migraine pain. These changes are likely based on gene expression of components like nNOS and CGRP receptor proteins and may therefore be very slow. Hence a therapeutic approach making use of these mechanisms can be expected to be successful rather in a prophylactic manner than in an acute intervention. To test hypotheses of such long-term drug actions beyond cell cultures, there is a need for new models that represent the complexity of expression changes in the TG and the respective effects in peripheral and central trigeminovascular tissues.

The chronic inhibition of 5-HT_{2B} receptors may be prophylactic in migraine

Antagonists of the 5-HT_{2B} receptor, methysergide, eyproheptadine and pizotifen, are effective in the prophylactic treatment of migraine (34,68), whereas ketanserin, which lacks affinity for 5-HT_{2B} or 5-HT_{2C} receptors, has no such prophylactic effect. The above substances act not only on 5-HT_{2B} but also on 5-HT_{2C} receptors, leaving the question open which of the receptor subtypes is responsible for the prophylactic effect.

Prophylactic migraine drugs, e.g. methysergide, can significantly reduce migraine frequency but this compound must be taken for a longer period (three to four weeks)

to achieve a therapeutic effect (69). Accordingly, in an animal model in rats, Saito et al. found that chronic but not acute treatment with methysergide inhibited PPE in the dura mater (70). The authors speculated that the difference between acute and chronic administration is the accumulation of the active metabolite, methylergotamine, as the actually effective drug.

Schaerlinger et al. found in an in vitro system (transfected human 5-HT_{2B/2C} in LMTK⁻ cells) that the long-term use of dihydroergotamine (DHE) leads to desensitization of 5-HT_{2B} receptors but not 5-HT_{2C} receptors (71). In transfected Chinese Hamster Ovary (CHO)-K1 cells, among all 5-HT₂ receptor subtypes, the 5-HT_{2B} receptors underwent the highest degree of desensitization to chronic 5-HT exposure (72). Moskowitz postulated in 1992 that methysergide and its metabolite methylergotamine inhibit the release of CGRP from perivascular sensory nerves (73). Though vasodilatation is no longer regarded as the key mechanism in migraine pain generation, limiting CGRP release, as it is achieved by 5-HT_{1B/D} agonists (triptans), is closely associated with an antimigraine effect. In a recent study for the first time mice could be made sensitive to 5-HT_{2B} receptor agonists by chronic hypoxia, a model that may demonstrate the potential importance of this receptor in chronic migraine processes. The authors postulated that four weeks' hypoxia induced a shift from a “non-migraineur” to a “migraineur-like” state, which was shown to depend on chronic activation of 5-HT_{2B} receptors. The mechanism for this shift could not be explained by an increased expression of 5-HT_{2B} receptors or other proteins involved. Learning more about the underlying cellular mechanisms of this phenomenon is demanding and could explain why the exclusively chronic treatment with 5-HT_{2B} receptor antagonists leads to a reduction of migraine attacks (38).

In conclusion, in search of a preventive antimigraine drug based on 5-HT₂ receptor inhibition, it is desirable to find a highly selective 5-HT_{2B} receptor antagonist that readily binds to the receptor and does not depend on an effective metabolite generated in the body. Second, such a receptor antagonist should have no or minimal activity at the 5-HT_{2C} receptor to reduce central side effects, e.g. psychedelic and hallucinogenic actions (74). Clinical studies are awaited to prove this concept.

Article highlights

- 5-HT_{2B} receptors are expressed at a low rate in a variety of tissues including blood vessels of the cranial dura and in the trigeminal ganglion.
- Activation of 5-HT_{2B} receptors causing neurogenic inflammation via nitric oxide synthesis indicates their possible involvement in migraine pathophysiology.
- Inhibition of 5-HT_{2B} receptors may contribute to a prophylactic effect in migraine.

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References

- Sicuteri F, Testi A and Anselmi B. Biochemical investigations in headache: Increase in hydroxyindole acid excretion during migraine attacks. *Int Arch Allergy Immunol* 1961; 19: 55–58.
- Kalkman HO. Is migraine prophylactic activity caused by 5-HT_{2B} or 5-HT_{2C} receptor blockade? *Life Sci* 1994; 54: 641–644.
- Panconesi A and Sicuteri R. Headache induced by serotonergic agonists—a key to the interpretation of migraine pathogenesis? *Cephalalgia* 1997; 17: 3–14.
- Anthony M, Lord GD and Lance JW. Controlled trials of cimetidine in migraine and cluster headache. *Headache* 1978; 18: 261–264.
- Somerville BW. Platelet-bound and free serotonin levels in jugular and forearm venous blood during migraine. *Neurology* 1976; 26: 41–45.
- Ferrari MD. Migraine. *Lancet* 1998; 351: 1043–1051.
- Brewerton TD, Murphy DL, Mueller EA, et al. Induction of migrainelike headaches by the serotonin agonist m-chlorophenylpiperazine. *Clin Pharmacol Ther* 1988; 43: 605–609.
- Schmitz B, Ullmer C, Segelcke D, et al. BF-1—A novel selective 5-HT_{2B} receptor antagonist blocking neurogenic dural plasma protein extravasation in guinea pigs. *Eur J Pharmacol* 2015; 751: 73–80.
- Johnson KW, Nelson DL, Dieckman DK, et al. Neurogenic dural protein extravasation induced by meta-chlorophenylpiperazine (mCPP) involves nitric oxide and 5-HT_{2B} receptor activation. *Cephalalgia* 2003; 23: 117–123.
- Martin RS and Martin GR. Investigations into migraine pathogenesis: Time course for effects of m-CPP, BW723C86 or glyceryl trinitrate on appearance of Fos-like immunoreactivity in rat trigeminal nucleus caudalis (TNC). *Cephalalgia* 2001; 21: 46–52.
- Schmuck K, Ullmer C, Kalkman HO, et al. Activation of meningeal 5-HT_{2B} receptors: An early step in the generation of migraine headache? *Eur J Neurosci* 1996; 8: 959–967.
- Vane JR. The relative activities of some tryptamine analogues on the isolated rat stomach strip preparation. *Br J Pharmacol Chemother* 1959; 14: 87–98.
- Foguet M, Hoyer D, Pardo LA, et al. Cloning and functional characterization of the rat stomach fundus serotonin receptor. *EMBO J* 1992; 11: 3481–3487.
- Foguet M, Nguyen H, Le H, et al. Structure of the mouse 5-HT_{1C}, 5-HT₂ and stomach fundus serotonin receptor genes. *Neuroreport* 1992; 3: 345–348.
- Schmuck K, Ullmer C, Engels P, et al. Cloning and functional characterization of the human 5-HT_{2B} serotonin receptor. *FEBS Lett* 1994; 342: 85–90.
- Kursar JD, Nelson DL, Wainscott DB, et al. Molecular cloning, functional expression, and mRNA tissue distribution of the human 5-hydroxytryptamine_{2B} receptor. *Mol Pharmacol* 1994; 46: 227–234.
- Barnes NM and Sharp T. A review of central 5-HT receptors and their function. *Neuropharmacology* 1999; 38: 1083–1152.
- Bonhaus DW, Bach C, DeSouza A, et al. The pharmacology and distribution of human 5-hydroxytryptamine_{2B} (5-HT_{2B}) receptor gene products: Comparison with 5-HT_{2A} and 5-HT_{2C} receptors. *Br J Pharmacol* 1995; 115: 622–628.
- Launay J, Hervé P, Peoc'h K, et al. Function of the serotonin 5-hydroxytryptamine 2B receptor in pulmonary hypertension. *Nat Med* 2002; 8: 1129–1135.
- Duxon MS, Flanigan TP, Reavley AC, et al. Evidence for expression of the 5-hydroxytryptamine_{2B} receptor protein in the rat central nervous system. *Neuroscience* 1997; 76: 323–329.
- Murray KC, Stephens MJ, Ballou EW, et al. Motoneuron excitability and muscle spasms are regulated by 5-HT_{2B} and 5-HT_{2C} receptor activity. *J Neurophysiol* 2011; 105: 731–748.
- Nicholson R, Small J, Dixon AK, et al. Serotonin receptor mRNA expression in rat dorsal root ganglion neurons. *Neurosci Lett* 2003; 337: 119–122.
- Lin S, Chang W, Lin C, et al. Serotonin receptor 5-HT_{2B} mediates serotonin-induced mechanical hyperalgesia. *J Neurosci* 2011; 31: 1410–1418.
- Pineda-Farias JB, Velázquez-Lagunas I, Barragán-Iglesias P, et al. 5-HT_{2B} receptor antagonists reduce nerve injury-induced tactile allodynia and expression of 5-HT_{2B} receptors. *Drug Dev Res*. Epub ahead of print 25 January 2015. DOI: 10.1002/ddr.21238.
- Florian JA and Watts SW. Integration of mitogen-activated protein kinase activation in vascular 5-hydroxytryptamine_{2A} receptor signal transduction. *J Pharmacol Exp Ther* 1998; 284: 346–355.
- Manivet P, Mouillet-Richard S, Callebert J, et al. PDZ-dependent activation of nitric-oxide synthases by the serotonin 2B receptor. *J Biol Chem* 2000; 275: 9324–9331.
- Cox DA and Cohen ML. 5-Hydroxytryptamine_{2B} receptor signaling in rat stomach fundus: Role of voltage-dependent calcium channels, intracellular calcium release and protein kinase C. *J Pharmacol Exp Ther* 1995; 272: 143–150.
- Richardson JD and Vasko MR. Cellular mechanisms of neurogenic inflammation. *J Pharmacol Exp Ther* 2002; 302: 839–845.
- Hamik A and Peroutka SJ. 1-(m-chlorophenyl)piperazine (mCPP) interactions with neurotransmitter receptors in the human brain. *Biol Psychiatry* 1989; 25: 569–575.
- Bonhaus DW, Flippin LA, Greenhouse RJ, et al. RS-127445: A selective, high affinity, orally bioavailable

- 5-HT_{2B} receptor antagonist. *Br J Pharmacol* 1999; 127: 1075–1082.
31. Bonhaus DW, Chang LK, Cao Z, et al. RS-127445, a selective 5-HT_{2B} receptor antagonist, blocks a mCPP induced plasma protein extravasation in dura mater and capsaicin-evoked c-fos expression in trigeminal nucleus caudalis. In: Olesen J, Goadsby PJ (eds) *Cluster headache and related headaches*. New York: Oxford University Press, 1999, pp.278–286.
 32. Mylecharane EJ. 5-HT₂ receptor antagonists and migraine therapy. *J Neurol* 1991; 238(Suppl. 1): S45–S52.
 33. Speight TM and Avery GS. Pizotifen (BC-105): A review of its pharmacological properties and its therapeutic efficacy in vascular headaches. *Drugs* 1972; 3: 159–203.
 34. Fozard JR. The 5-hydroxytryptamine-nitric oxide connection: The key link in the initiation of migraine? *Arch Int Pharmacodyn Ther* 1995; 329: 111–119.
 35. McDonald DM, Thurston G and Baluk P. Endothelial gaps as sites for plasma leakage in inflammation. *Microcirculation* 1999; 6: 7–22.
 36. Williamson DJ and Hargreaves RJ. Neurogenic inflammation in the context of migraine. *Microsc Res Tech* 2001; 53: 167–178.
 37. Dimitriadou V, Buzzi MG, Theoharides TC, et al. Ultrastructural evidence for neurogenically mediated changes in blood vessels of the rat dura mater and tongue following antidromic trigeminal stimulation. *Neuroscience* 1992; 48: 187–203.
 38. Hunfeld A, Segelcke D, Bäcker I, et al. Hypoxia facilitates neurogenic dural plasma protein extravasation in mice: A novel animal model for migraine pathophysiology. *Sci Rep* 2015; 5: 17845.
 39. Ghabriel MN, Lu MX, Leigh C, et al. Substance P-induced enhanced permeability of dura mater microvessels is accompanied by pronounced ultrastructural changes, but is not dependent on the density of endothelial cell anionic sites. *Acta Neuropathol* 1999; 97: 297–305.
 40. Goldstein DJ, Offen WW, Klein EG, et al. Lanepitant, an NK-1 antagonist, in migraine prevention. *Cephalalgia* 2001; 21: 102–106.
 41. Goldstein DJ, Wang O, Saper JR, et al. Ineffectiveness of neurokinin-1 antagonist in acute migraine: A crossover study. *Cephalalgia* 1997; 17: 785–790.
 42. Williamson DJ, Hargreaves RJ, Hill RG, et al. Sumatriptan inhibits neurogenic vasodilation of dural blood vessels in the anaesthetized rat—intravital microscope studies. *Cephalalgia* 1997; 17: 525–531.
 43. Williamson DJ, Shephard SL, Hill RG, et al. The novel anti-migraine agent rizatriptan inhibits neurogenic dural vasodilation and extravasation. *Eur J Pharmacol* 1997; 328: 61–64.
 44. Dimitriadou V, Lambracht-Hall M, Reichler J, et al. Histological and ultrastructural characteristics of rat brain perivascular mast cells stimulated with compound 48/80 and carbachol. *Neuroscience* 1990; 39: 209–224.
 45. Gupta S, Nahas SJ and Peterlin BL. Chemical mediators of migraine: Preclinical and clinical observations. *Headache* 2011; 51: 1029–1045.
 46. Levy D, Burstein R, Kainz V, et al. Mast cell degranulation activates a pain pathway underlying migraine headache. *Pain* 2007; 130: 166–176.
 47. Schwenger N, Dux M, de Col R, et al. Interaction of calcitonin gene-related peptide, nitric oxide and histamine release in neurogenic blood flow and afferent activation in the rat cranial dura mater. *Cephalalgia* 2007; 27: 481–491.
 48. Toda N. Mechanism underlying responses to histamine of isolated monkey and human cerebral arteries. *Am J Physiol* 1990; 258: H311–H317.
 49. Ottosson AL, Jansen I, Langemark M, et al. Histamine receptors in the isolated human middle meningeal artery. *A comparison with cerebral and temporal arteries*. *Cephalalgia* 1991; 11: 183–188.
 50. Russell D. Cluster headache: Trial of a combined histamine H₁ and H₂ antagonist treatment. *J Neurol Neurosurg Psychiatr* 1979; 42: 668–669.
 51. Fozard JR. The animal pharmacology of drugs used in the treatment of migraine. *J Pharm Pharmacol* 1975; 27: 297–321.
 52. Brian JE, Faraci FM and Heistad DD. Recent insights into the regulation of cerebral circulation. *Clin Exp Pharmacol Physiol* 1996; 23: 449–457.
 53. Messlinger K, Lennerz JK, Eberhardt M, et al. CGRP and NO in the trigeminal system: Mechanisms and role in headache generation. *Headache* 2012; 52: 1411–1427.
 54. Singh S, Padovani D, Leslie RA, et al. Relative contributions of cystathionine beta-synthase and gamma-cystathionase to H₂S biogenesis via alternative trans-sulfuration reactions. *J Biol Chem* 2009; 284: 22457–22466.
 55. Kunkler PE, Ballard CJ, Oxford GS, et al. TRPA1 receptors mediate environmental irritant-induced meningeal vasodilatation. *Pain* 2011; 152: 38–44.
 56. Eberhardt M, Dux M, Namer B, et al. H₂S and NO cooperatively regulate vascular tone by activating a neuroendocrine HNO-TRPA1-CGRP signalling pathway. *Nat Commun* 2014; 5: 4381.
 57. Edvinsson L, Ekman R, Jansen I, et al. Calcitonin gene-related peptide and cerebral blood vessels: Distribution and vasomotor effects. *J Cereb Blood Flow Metab* 1987; 7: 720–728.
 58. Dux M, Will C, Vogler B, et al. Meningeal blood flow is controlled by H₂ S-NO crosstalk activating HNO-TRPA1-CGRP signalling. *Br J Pharmacol* 2016; 173: 431–445.
 59. Bellamy J, Bowen EJ, Russo AF, et al. Nitric oxide regulation of calcitonin gene-related peptide gene expression in rat trigeminal ganglia neurons. *Eur J Neurosci* 2006; 23: 2057–2066.
 60. Li J, Vause CV and Durham PL. Calcitonin gene-related peptide stimulation of nitric oxide synthesis and release from trigeminal ganglion glial cells. *Brain Res* 2008; 1196: 22–32.
 61. Kleschyov AL, Oelze M, Daiber A, et al. Does nitric oxide mediate the vasodilator activity of nitroglycerin? *Circ Res* 2003; 93: e104–e112.
 62. Olesen J, Thomsen LL, Lassen LH, et al. The nitric oxide hypothesis of migraine and other vascular headaches. *Cephalalgia* 1995; 15: 94–100.

63. Dieterle A, Fischer, Michael JM, et al. Increase in CGRP- and nNOS-immunoreactive neurons in the rat trigeminal ganglion after infusion of an NO donor. *Cephalalgia* 2011; 31: 31–42.
64. Seiler K, Nusser JI, Lennerz JK, et al. Changes in calcitonin gene-related peptide (CGRP) receptor component and nitric oxide receptor (sGC) immunoreactivity in rat trigeminal ganglion following glyceroltrinitrate pretreatment. *J Headache Pain* 2013; 14: 74.
65. Storer RJ, Akerman S and Goadsby PJ. Calcitonin gene-related peptide (CGRP) modulates nociceptive trigemino-vascular transmission in the cat. *Br J Pharmacol* 2004; 142: 1171–1181.
66. Schlechtweg PM, Röder J, Fischer MJ, et al. Increase in NADPH-diaphorase-positive and neuronal NO synthase immunoreactive neurons in the rat spinal trigeminal nucleus following infusion of a NO donor—evidence for a feed-forward process in NO production involved in trigeminal nociception. *Cephalalgia* 2009; 29: 566–579.
67. Xu L, Mabuchi T, Katano T, et al. Nitric oxide (NO) serves as a retrograde messenger to activate neuronal NO synthase in the spinal cord via NMDA receptors. *Nitric Oxide* 2007; 17: 18–24.
68. Fozard JR and Kalkman HO. 5-Hydroxytryptamine (5-HT) and the initiation of migraine: New perspectives. *Naunyn Schmiedebergs Arch Pharmacol* 1994; 350: 225–229.
69. Lance JW, Anthony M and Somerville B. Comparative trial of serotonin antagonists in the management of migraine. *Br Med J* 1970; 2: 327–330.
70. Saito K, Markowitz S and Moskowitz MA. Ergot alkaloids block neurogenic extravasation in dura mater: Proposed action in vascular headaches. *Ann Neurol* 1988; 24: 732–737.
71. Schaerlinger B, Hickel P, Etienne N, et al. Agonist actions of dihydroergotamine at 5-HT_{2B} and 5-HT_{2C} receptors and their possible relevance to antimigraine efficacy. *Br J Pharmacol* 2003; 140: 277–284.
72. Porter RH, Malcolm CS, Allen NH, et al. Agonist-induced functional desensitization of recombinant human 5-HT₂ receptors expressed in CHO-K1 cells. *Biochem Pharmacol* 2001; 62: 431–438.
73. Moskowitz MA. Neurogenic versus vascular mechanisms of sumatriptan and ergot alkaloids in migraine. *Trends Pharmacol Sci* 1992; 13: 307–311.
74. Abramson HA and Rolo A. Lysergic acid diethylamide (LSD-25). 38. Comparison with action of methysergide and psilocybin on test subjects. *J Asthma Res* 1965; 3: 81–96.